

# ELECTROPHORETIC APPARATUS FOR THE QUANTITATIVE ELUTION OF PROTEINS OR POLYPEPTIDES FROM A GEL

## BACKGROUND OF THE INVENTION

The present invention relates to an apparatus for the quantitative elution of proteins or polypeptides from a gel by means of electrophoresis.

Protein mixtures, such as viruses, and isolated biological membranes, etc. (separated according to SDS (sodium dodecyl sulfate)/polyacrylamide gel electrophoresis (see e.g. Laemmli, *Nature* Vol. 227, pp. 680 to 685, 1970; and Thomas and Kornberg, *Proc. Nat. Acad. Sci. U.S.A.* Vol. 72, pp. 2226 to 2630, 1975) are represented as individual bands in a plate gel. These bands correspond to individual proteins having the corresponding molecular weight. In order to utilize the thus separated proteins, or polypeptides, for other purposes, the bands are separated from the plate gel and collected. The protein retained in the network of the polyacrylamide is electrophoretically eluted. Generally a running buffer such as 5 mM tris-glycin, pH 8.6 0.2% SDS and 0.5% mercaptoethanol is used as the elution buffer.

## SUMMARY OF THE INVENTION

It is an object of the present invention to provide an apparatus with which isolation of even larger quantities of electrophoretically separated proteins or polypeptides is possible in a desired concentration.

The above and other objects are accomplished in accordance with the invention wherein an apparatus is provided for the quantitative elution of proteins or polypeptides from a gel by means of electrophoresis. The apparatus includes an upper electrode chamber for holding a buffer solution containing the gel from which the proteins or polypeptides are to be eluted. An upper electrode is mounted in the upper electrode chamber. A lower electrode chamber for holding buffer solution is disposed beneath the upper electrode chamber. A lower electrode is mounted in the lower electrode chamber. A septum separates the upper electrode chamber from the lower electrode chamber. The septum is provided with a connecting passage for connecting the upper and lower chambers. A collecting capsule for collecting the proteins or polypeptides is disposed at the end of the connecting passage in the lower electrode chamber, and is adapted to be suspended in the buffer solution which is to be held in the lower chamber.

In operating the apparatus, the application of a voltage potential between the upper and lower electrodes causes the proteins or polypeptides to be eluted from the gel in the upper chamber and collected in the collecting capsule.

It is to be understood that both the foregoing general description and the following detailed description are exemplary, but are not restrictive of the invention.

## BRIEF DESCRIPTION OF THE DRAWING

The sole FIGURE is a cross-sectional view of an elution apparatus according to the invention.

## DESCRIPTION OF THE PREFERRED EMBODIMENT

Referring now to the drawing, there is shown an elution apparatus which includes an upper electrode chamber 4 which is to contain a buffer solution and a lower electrode chamber 10 which is to contain the

same buffer solution. The individual proteins or polypeptides separated and collected from the plate gel which are to be eluted, are placed in the buffer solution in upper electrode chamber 4.

Chambers 4 and 10, both of which can be of cylindrical design, are placed one on top of the other and are separated from each other by a septum 30. Septum 30 is provided with a connecting passage 13 which connects upper electrode chamber 4 with lower electrode chamber 10.

Chamber 4 is formed by a side wall 32, a bottom 16 and a removable top 3. Bottom 16 generally comprises the upper surface of septum 30 which preferably is integral (in one piece) with side wall 32. A negative electrode 1 is provided in top 3.

Chamber 10 is formed by a side wall 34, a bottom 12 and a top 36 which is the lower surface of septum 30. A positive electrode 11 is provided in bottom 12 of chamber 10.

Electrodes 1 and 11 are illustrated with a planar shape, but could have an annular or other suitable shape. Both electrodes 1 and 11 are disposed perpendicular to the longitudinal axis (axis of rotation) of chambers 4 and 10, which also passes through connecting passage 13.

Upper electrode 1 is fastened to a mount 14 which passes through top 3. A guide ring 15 is attached to the upper surface of top 3 and is provided with a thumb screw 2. Mount 14 passes through guide ring 15 and can be set at any desired height by adjusting thumb screw 2. Thus, upper electrode 1 can be adjusted in height with respect to the surface (not shown) of the gel-buffer solution disposed in upper electrode chamber 4. The height at which electrode 1 is set depends on the fill level of the gel-buffer solution in chamber 1 and the electrical voltage between electrodes 1 and 11. The level of the elution buffer in the upper electrode chamber 4 should be 1.5 to 2 cm below the upper rim of the chamber. The electrode 1 of the upper chamber should dip about 1 cm into the buffer solution. The lower electrode chamber 10 should be filled with buffer up to the level of the counter ring 8.

The electrical voltage between the electrodes 1 and 11 should not exceed 100 V.

Bottom 16 of upper electrode chamber 4 is preferably constructed in the form of a funnel having a downwardly narrowing conical inclined surface 38, a funnel neck 40 which meets with inclined surface 38, and a seat 42 which meets with funnel neck 40. The lowest point of bottom 16 is at seat 42 and is joined to connecting passage 13. A small tube 7 having an upper end 44 and a lower end 19 is disposed in passage 13 and extends into the area defined by funnel neck 40 and seat 42. Tube 7 is provided with an abutment 17 which meets a corresponding abutment 46 in passage 13. Abutments 17 and 46 together define the extent to which tube 7 can be inserted from lower electrode chamber 10 into connecting passage 13 and the area defined by neck 40 and seat 42. A radial packing ring 6 is provided at neck 40 and seals tube 7 relative to upper electrode chamber 4. Sinter plates 5 may cover upper end 44 and lower end 19 of tube 7. The radial packing ring 6 is a oil seal or rubber O-ring. Glass sinter plates 5 of 150  $\mu$ m to 50  $\mu$ m pore sizes can be used.

Lower end 19 of tube 7 has a frustoconical shape. A dialysis capsule 9 is clamped onto end 19 by a counter cone ring 8.